### Development of TOF-MS from intellectual curiosity to practical technique

Marvin Vestal SimulTOF Systems The Jim Waters Symposium at PittCon March 2, 2014

### A Brief History of Time (of flight) with apologies to Stephen Hawking

- 1946 W.E Stephens, Phys. Rev. 69,641
  - "Advances in electronics seem to make practical a type of mass spectrometer in which microsecond pulses of ions are selected every millisecond from an ordinary low-voltage ion source. In travelling down the vacuum tube, ions of different M/e have different velocities and consequently separate into groups spread out in space. ... This type of mass spectrometer should offer many advantages over present types. The response time should be limited only by the repetition rate (milliseconds)... Magnets and stabilization equipment would be eliminated. Resolution would not be limited by smallness of slits or alignment. Such a mass spectrometer should be well suited for composition control, rapid analysis, and portable use."

### Brief History of Time (of flight)

- 1948 Cameron & Eggers, *Rev. Sci. Instr.* 19, 605.
  - First working TOF
- 1953 Wiley & McLaren, Rev. Sci. Instr. 26, 1150
  - First practical TOF. Energy & Time Lag Focusing.
- 1959 Gohlke, Anal. Chem. **31**, 535

- GC-TOF

- 1963 Vestal & Wharhaftig, ASMS, 358.
  - Coincidence TOF, first ion counting TDC
- 1973 Mamyrin et al, Sov. Phys. JETP 37, 45.
  - Reflectron, higher resolution
- 1974 Macfarlane et al, Biochem. Biophys. Res. Comm. 60, 616
  <sup>252</sup>Cf Plasma desorption. Proteins Fly!!!!

### Brief History of Time (of flight)

- 1988 Karas & Hillenkamp, Anal. Chem. 60, 2299.
  - MALDI Really big proteins fly!!!
- 1991 Dodenov et al, 12th Int. MS Conf.
  - O-TOF Electrospray works with TOF
- 1993 Kaufman, Spengler & Lutzenkirchen, RCM 7, 902.
  - Post-source decay MALDI
- 1994 Brown & Lennon, Sunriver, p63.
  - Delayed extraction MALDI Makes MALDI-TOF routine
- **1996** Morris et al, *RCM* **10**, 889.
  - Q-TOF
- 2001
  - TOF-TOF

Early History of MALDI-TOF



#### Karras & Hillenkamp 1988



Beavis & Chait 1989





Initial velocity of ions from MALDI is high and independent of mass. This is the major contribution to time dispersion in static MALDI.



Delayed extraction source plus 2-stage reflector Makes high resolution MALDI practical

Table 1.	Effective flight paths for the employed in this work	e mass analyzers
Analyzer	Linear (m)	Reflector (m)
RP	1.3	2.0
EL	2.0	3.0
XL	4.2	6.6



Figure 3. Comparison of the resolution and signal-to-noise (S/N) ratio obtained for the molecular ion region of angiotensin I (MH<sup>+</sup>, monoisotopic m/z = 1296.68) in three different operating modes (static linear, delayed extraction linear and delayed extraction reflector) as a function of the analyzer geometry (RP, EL, XL) as given in Table 1. Matrix:  $\alpha$ -cyano-4-hydroxycinnamic acid.

Contributions to relative peak width,  $\Delta m/m$ 

Initial position,  $\delta x$ :  $R_{s1} = [(D_v - D_s)/D_e](\delta x/d_0 y)$ 

Initial velocity,  $\delta v_0$ :  $R_{v1} = (4d_0y/D_e)(\delta v_0/v_n)[(1-(m/m^*)^{1/2}]$   $R_{v2} = 2[2d_0y/(D_v-D_s)]^2 (\delta v_0/v_n)^2$  $R_{v3} = 2[2d_0y/(D_v-D_s)]^3 (\delta v_0/v_n)^3$ 

Time error,  $\delta t$ : $R_t = 2\delta t/t = 2\delta tv_n/D_e$ Trajectory error,  $\delta L$ : $R_L = 2\delta L/D_e$ Not small!!!Voltage error,  $\delta V$ : $R_V = \delta V/V$ 

Resolving power:

 $R^{-1} = [R_{s1}^{2} + R_{v}^{2} + R_{t}^{2} + R_{L}^{2} + R_{v}^{2}]^{-1/2}$ 

Contributions to relative peak width,  $\Delta m/m$ , with 1<sup>st</sup> and 2<sup>nd</sup> order velocity focusing (size matters) Initial position,  $\delta x$ :  $R_{s1} = [2/K](\delta x/D_e)$ 

Initial velocity,  $\delta v_0$ :  $R_{v1} = (4d_0y/D_e)(\delta v_0/v_n)[(1-(m/m^*)^{1/2}]$   $R_{v2} = 2[K]^2 (\delta v_0/v_n)^2 = 0$  $R_{v3} = 2[K]^3 (\delta v_0/v_n)^3$ 

Time error,  $\delta t$ :  $R_t = 2\delta t/t = 2\delta t v_n / D_e$ 

Trajectory error,  $\delta L$ :  $R_1 = 2\delta L/D_e$ 

Voltage error,  $\delta V$ :  $R_v = \delta V/V$ 

**Resolving power:** 

$$R^{-1} = [R_{s1}^{2} + R_{v}^{2} + R_{t}^{2} + R_{L}^{2} + R_{v}^{2}]^{-1/2}$$

 $\delta v_0$ =400 m/s,  $\delta x_0$ =0.01 mm,  $\delta t$ =0.75 ns



Maximum resolving power at focus mass under optimized conditions.

Does not include contributions from trajectory error, HV noise, and collisions

----- Group:1 Spot:277 Shots:500 Peaks:1,182 File:L13

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#### 6 m single mirror 14 m effective

Source is on first floor and mirror on second floor







Calibration Equation:  $m^{1/2}=D_0 + D_1 t[1+D_2 t+D_3 t^2]$ Determine coefficients by least square fit to multiple Peaks covering broad mass range



RMS error for 10 peaks in spectrum of tryptic digest of BSA for all 675 spots on a 102x108 mm sample plate with automatic 2-point internal Calibration for 3.2 m analyzer



JEOL Spiraltron 17m



#### Mass resolving power



# **High Resolution MALDI**

- Resolving power >20,000 for peptides and small molecules is routine
- Resolving Power approaching 100,000 has been demonstrated with long flight distances
- RMS Mass error 1-5 ppm across the plate over the full mass range with single peak automatic calibration is routine
- MALDI ions often metastable thus longer flight time imply lower sensitivity particularly for larger, more fragile ions
- Sensitivity generally limited by chemical noise, thus higher resolving power may yield higher sensitivity

For Linear Analyzer (size matters more) Initial position,  $\delta x$ :  $R_{s1} = 2[(D_v - D_s)/2d_0y](\delta x/D_e)$ 

Initial velocity,  $\delta v_0$ :

$$R_{v1} = (4d_0y/D_e)(\delta v_0/v_n)[(1-(m/m^*)^{1/2}] R_{v2} = 2[2d_0y/(D_v-D_s)]^2 (\delta v_0/v_n)^2 = 0 R_{v3} = 2[2d_0y/(D_v-D_s)]^3 (\delta v_0/v_n)^3$$

Time error,  $\delta t$ :  $R_t = 2\delta t/t = 2\delta t v_n/D_e$ 

Trajectory error,  $\delta L$ :  $R_L = 2\delta L/D_e$ 

Voltage error,  $\delta V$ :  $R_v = \delta V/V$ 

Resolving power:

$$R^{-1} = [R_{s1}^{2} + R_{v}^{2} + R_{t}^{2} + R_{L}^{2} + R_{v}^{2}]^{-1/2}$$

#### Sensitivity, Dynamic Range, and Reproducibility are Key Metrics



Maximum Resolving Power for Linear Analyzer.



Linear @10 kV -- Higher Resolving Power than the old XL Reflector

#### Photonis detector



Potential diagram for linear detector

### Typical single ion pulse with fast scintillator





Fig. 9 BSA (1 picomole/ $\mu$ l) in sinnipinic acid matrix 10,000 laser shots in 2 s.

### **Conventional Wisdom**

# • MALDI

• is Not

Quantitive



4 spectra 50 shots ea. On one sample spot Intensity varies by factor of 20 @10 Hz requires 20 s

Your Old MALDI

### Can MALDI be Quantitative????

#### Data Acquisition, One Example

Data is acquired over mass range 1000-20,000 Da using 2 ns bins 2 kHz laser rate 50 shots averaged per spectrum 1 mm/s snake raster for 5 passes at 500 mm over 3 mm dia spot Total travel 12 mm in 12 s generating 480 spectra with 50 µm resolution Spectra with no significant peaks are not saved Only 10% of sample used, 240,000 total laser shots possible

#### **Data Processing**

Spot Average Baseline Correction Smooth Calibrate and Detect Peaks Normalize Bin by mass rather than time Quantify Report





Spot 176

Group:1 Number:220J4 Shots:14,050 Peaks:161 —— Group:1 Number:221J5 Shots:15,750 Peaks:165



Spectra from 4 different sample spots super imposed

# Linear MALDI

- Resolving power is mass dependent
  - 5000-8000 over limited range at low mass
  - 500-1000 over wide range limited by width of isotope distribution
- Each spot will yield up to 200,000 shots without degrading resolving power or accuracy and giving dynamic range limited only by chemical noise
- Mass error <30 ppm across the plate over the full mass range with single peak automatic calibration
- Dynamic range up to100,000
- High laser rate and data processing makes MALDI quantitative







----- Group:1 Spot:487 Shots:529,000 Peaks:199 PM:1296.685





# Status of TOF-TOF

- Trade-off between high resolution precursor selection and high sensitivity for fragments
- 5 kHz laser makes LC-MALDI-MS-MS practical
- Multiplexing up to 10 precursors/laser shot practical for MRM measurements
- Both low energy fragmentation (psd) and high energy(cid) are practical
- Still not widely accepted relative to ESI with traps

# Applications

- Pathogen Identification
- Cancer typing directly from serum, tissue extracts, and other bodily fluids
- Tissue imaging
  - Proteins for cancer typing
  - Small molecules for drug disposition
- Biomarker Identification and Validation
  - Mass Spec Immunoassay
  - Peptide quantitation (SISCAPA and others)
- Clinical assays of biomarkers for diagnosis and treatment
- Protein array reader